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Total Synthesis of Trehalase Inhibitor Salbostatin

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Abstract: Trehalase inhibitor salbostatin has been completely synthesized by coupling of 1,5:2,3-dianhydro-D-mannitol and di-*O*-isopropylidene-α-valienamine followed by deprotection.

Very recently, new trehalase inhibitor salbostatin (1) was discovered as a novel metabolite of *Streptomyces albus*, ATCC 21838, and the structure has been established mainly on the basis of 1 H NMR spectroscopic data. Salbostatin inhibits trehalase from porcine kidneys with an inhibition constant $Ki = 1.8 \times 10^{-7}$ M, and possesses a very unique pseudo-disaccharide structure composed of 2-amino-1,5-anhydro-2-deoxy-D-glucitol to which an unsaturated 5a-carba-sugar, α -valienamine (2) residue, is attached by way of an imino bridge. Similar pseudo-disaccharidic glycosidase inhibitors containing 2, validoxylamine A^3 (3) and methyl α -acarviosin (4), a core component of acarbose, have been known so far. The former is a potent trehalase inhibitor and its dihydro derivative more likely mimicking substrate α , α '-trehalose-structure exhibits also high inhibitory activity.

We have been studying a structure-inhibitory activity relationship of this type of inhibitors⁵ and, especially, our interests now have been focused on trehalase inhibitors such as 3 and trehazolin³,6. In this paper, convenient total synthesis of salbostatin has been attempted in order both to confirm the structure proposed and to elaborate a general method for the preparation of salbostatin analogues. The method involves coupling of an anhydro sugar between versatile carba-sugar donor 2,3:4,6-di-O-isopropylidene- α -valienamine⁷ (7).

As anhydro sugar acceptor, we chose the unprotected 1,5:2,3-dianhydro-D-mannitol (6), which was readily derived (95%) by hydrogenolysis (10% Pd/C) of the known corresponding 4,6-O-benzylidene derivative⁸ (5). Removal of the benzylidene group of 5 seemed to enhance reactivity of the 2,3-epoxide and, hopefully, to improve desired regioselectivity of its cleavage by the bulky amine, owing to relief from the structural rigidity. Coupling of a slight excess of 6 (1.3 molar equivalent) and 7 was thus carried out conventionally⁹ in 2-propanol in a sealed tube for 4 days at 120°C. TLC showed as had been expected a formation of two coupling products. Chromatography of the mixture on a silica gel column with butanone–toluene (3:1, v/v) as an eluent afforded the diequatorial-opening product 8 (58%) and the diaxial-opening product 9

(25%), the structures of which were tentatively assigned as depicted in Scheme on the basis of the 1 H NMR spectra. The spectrum (270 MHz, CDCl₃) of 8 was well resolved, being amenable to a first-order analysis. Thus, the coupled signals due to 1,1-H and 2-H appeared as a doublet of doublets (6 4.08, J = 4.8 and 11.0 Hz), a triplet (6 3.19, J = 11.0 and 11.0 Hz), and a doublet of doublets of doublets (6 2.80, J = 4.8, 9.2, and 11.0 Hz), respectively, supporting an 1,5-anhydroglucitol structure of the sugar moiety. Preferential attack of the amine at C-2 may be due to the C-1 position being unsubstituted, and might be further rationalized by assuming that 6-hydroxyl group of 6 involves in stabilizing the favorable transition state that leads to diequatorial product. 7,10 O-Deisopropylidenation of 8 was effected in aqueous 70% acetic acid for 1 h at 60°C to give, after purification by a column of Dowex 50W-X2 (H+) resin with N aqueous ammonia, salbostatin 1 in 97% yield. The 13 C and 1 H NMR spectra 11 were in good accordance with those of an authentic sample. 2 The synthetic 1 was further characterized as the hepta-O-acetyl derivative 1a, the 1 H NMR spectrum 12 0 of which was shown to be superimposable on that reported. 2,13

The present synthesis constitutes the first total synthesis of the inhibitor salbostatin **1**, thereby confirming the structure proposed, and also provides one of the convenient methods for preparation of its analogues useful for elucidation of the structure-activity relationship.

References and Notes

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- 11. Data for salbostatin (1): $[\alpha]D^{25} + 130^{\circ}$ (c = 1.2, H₂O) [ref.² $[\alpha]D^{20} + 115^{\circ}$ (c = 1, H₂O)], ¹H NMR (270 MHz, $[D_6]DMSO-D_2O$, 4:1, v/v) δ 5.81 (d, $J_{1',2'} = 3.3$ Hz, 1 H, 2'-H), 4.05 and 3.99 (2 d, $J_{7'gem} = 14.4$ Hz, each 1 H, 7',7'-H), 3.91 (dd, $J_{1eq,2} = 4.6$, $J_{1gem} = 11.2$ Hz, 1 H, 1eq-H), 3.79 (d, $J_{4',5'} = 5.5$ Hz, 1 H, 4'-H), 3.69 (d, $J_{5,6a} = \sim 0$, $J_{6gem} = 12.1$ Hz, 1 H, 6a-H), 3.48 (dd,

- $J_{4',5'} = 5.5$, $J_{5',6'} = 8.8$ Hz, 1 H, 5'-H), 3.45 (dd, $J_{5,6b} = 5.9$, $J_{6gem} = 12.1$ Hz, 1 H, 6b-H), 3.41 (dd, $J_{1',6'} = 4.8$, $J_{5',6'} = 8.8$ Hz, 1 H, 6'-H), 3.23 (br dd, $J_{1',2'} = 3.3$, $J_{1',6'} = 4.8$ Hz, 1 H, 1'-H), 3.16–3.11 (m, 3 H, 3,4,5-H), 3.02 (dd, $J_{1ax,2} = 10.8$, $J_{1gem} = 11.2$ Hz, 1 H, 1ax-H), 2.66 (ddd, $J_{1ax,2} = 10.8$, $J_{1eq,2} = 4.6$, $J_{2,3} = 10.3$ Hz, 1 H, 2-H). I_{3}^{1} C-NMR (67.5 MHz, [D6]DMSO) I_{3}^{1} 140.38, 120.17, 81.67, 76.60, 72.90, 70.90, 70.47, 70.15, 68.97, 61.37, 61.25, 57.54, 52.76
- 12. Data for the hepta-O-acetyl derivative 1a: $[\alpha]D^{24} + 66.6^{\circ}$ (c = 0.75, CHCl3), ${}^{1}H$ NMR (270 MHz, CDCl3) δ 5.86 (br d, $J_{1',2'} = 4.8$ Hz, 1 H, 2'-H), 5.53 (d, $J_{4',5'} = 6.6$ Hz, 1 H, 4'-H), 5.48 (dd, $J_{4',5'} = 6.6$, $J_{5',6'} = 9.7$ Hz, 1 H, 5'-H), 4.97 (dd, $J_{1',6'} = 4.0$, $J_{5',6'} = 9.7$ Hz, 1 H, 6'-H), 4.95 (dd, $J_{3,4} = 9.3$, $J_{4,5} = 9.3$ Hz, 1 H, 4-H), 4.87 (dd, $J_{2,3} = J_{3,4} = 9.3$ Hz, 1 H, 3-H), 4.64 and 4.36 (2 d, $J_{7'}$ gem = 13.2 Hz, each 1 H, 7',7'-H), 4.21 (dd, $J_{5,6a} = 4.8$, $J_{6gem} = 12.5$ Hz, 1 H, 6a-H), 4.09 (dd, $J_{5,6b} = 2.2$, $J_{6gem} = 12.5$ Hz, 1 H, 6b-H), 4.08 (dd, $J_{1eq,2} = 4.8$, $J_{1gem} = 11.7$ Hz, 1 H, 1eq-H), 3.65 (br dd, $J_{1',2'} = 4.8$, $J_{1',6'} = 4.0$ Hz, 1 H, 1'-H), 3.54 (ddd, $J_{4,5} = 9.3$, $J_{5,6a} = 4.8$, $J_{5,6b} = 2.2$ Hz, 5-H), 3.14 (dd, $J_{1ax,2} = 11.0$, $J_{1gem} = 11.7$ Hz, 1 H, 1ax-H), 2.88 (ddd, $J_{1ax,2} = 11.0$, $J_{1eq,2} = 4.8$, $J_{2,3} = 9.3$ Hz, 1 H, 2-H), 2.10, 2.09, 2.06, 2.04, 2.03, and 2.02 (6 s, 3, 6, 3, 3, 3, and 3 H, 7 Ac).
- 13. We express our sincere thanks to Dr. L. Vértesy (Hoechst AG, Frankfurt am Main, FRG) for identification of the synthetic 1 and 1a with a natural compound and its derivative by comparison of their ¹NMR spectra.

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